Health Certificate No._____
(Valid only if USDA Veterinary Seal Appears Over the Certificate No.)

Health Certificate for Export of Bovine Embryos from the United States to New Zealand

1. Consignor: Name: Address:	2. Certificate reference number:
	3. Veterinary Authority: **USDA, APHIS**
	4. Import permit number:
5. Consignee: Name: Address:	6. Country of origin: **UNITED STATES**
	7. Country of destination: **NEW ZEALAND**
8. Approved embryo collection team: Name: Address:	9. Approval number of embryo collection team:
10. Place of shipment:	11. Date of departure:
12. Description of commodity: **BOVINE EMBRYOS**	13. Total number of embryos/straws:

DONOR (sire and dam) AND EMBRYO IDENTIFICATION

Name	Registration number	Breed	Date of birth	Collection date	Collection code	Straw ID	Number of embryos/ straws

Health Certificate No._____
(Valid only if USDA Veterinary Seal Appears Over the Certificate No.)

HEALTH CERTIFICATION

I,…….., an approved veterinarian authorized by APHIS certify, after due enquiry, that the embryos described above satisfy(ies) the following requirements:

Country freedom

1. For the last 12 months prior to embryo collection for export to New Zealand, the United States has been free of foot and mouth disease, Borna disease, Crimean Congo haemorrhagic fever, lumpy skin disease, Rift Valley fever, and contagious bovine pleuropneumonia, and these diseases are officially notifiable.

Donor eligibility

2. Donors that were imported into the United States have lived continuously in the United States for at least 60 days and in the herd of origin for at least 30 days prior to embryo collection for export to New Zealand.

3. Donors were resident in the embryo collection herd for at least 30 days prior to embryo collection for export to New Zealand.

Embryo collection team and herd approval requirements

4. At the time of embryo collection for export to New Zealand, the embryo collection team was approved by and registered with APHIS to collect, process, and store bovine embryos for export in accordance with the current recommendations of the OIE Code and the current manual of the International Embryo Transfer Society (IETS).

5. The approved embryo collection team veterinarian, under the supervision of APHIS, has knowledge of and authority over the embryo collection herd until completion of testing specified in this certificate.

Donor and herd health status

6. The donors were not resident in any establishment that is subject to quarantine restrictions, for at least the 60 days before the first embryo collection for the consignment to New Zealand until completion of the testing of the donors as required by this certificate.

7. Where a specific requirement for a risk organism is met by pre-collection testing, donors were isolated from other cattle not of an equivalent tested health status, from the time of the pre-collection test until completion of embryo collection for export to New Zealand.

8. On the day(s) of collection of the embryos, the donors were free from clinical evidence of infectious diseases transmissible in embryos, and the health status of the donors was monitored and recorded.

9. The semen used to produce the embryos in the consignment either:
    a. was imported directly from New Zealand or is eligible for export to New Zealand; OR
    b. was collected from a bull resident in a Certified Semen Services (CSS) participating herd; OR
    c. was collected and processed at a semen collection centre that fully complies with the current OIE Code chapter on collection and processing of bovine semen; OR
    d. where natural service or fresh semen was used, donor males were inspected and found free from clinical evidence of infectious diseases transmissible in semen, and were of an equivalent isolation and tested health status to the donor females.

Embryo collection, processing, storage and transport

10. All embryos in the consignment were fertilized *in vivo*, collected, washed, processed, traceability maintained, stored, and transported under the supervision of an approved embryo collection team veterinarian and in accordance with the recommendations in the OIE Code chapters on collection and processing of *in vivo* derived and micro-manipulated bovine embryos.

11. The embryos were treated with trypsin during the washing process as described in the IETS Manual. Each embryo had an intact zona pellucida and was examined over its entire surface at not less than 50X magnification and found to be free of adherent material.

12. Any micro-manipulation that causes a breach of the zona pellucida was done as per the procedures described in the OIE Code and IETS Manual and included specifications on the facilities used. Micro-manipulation was carried out on an embryo having an intact zona pellucida and was done subsequent to the last wash and examination of the embryo.

13. All biological products of animal origin used in the media and solutions for collection, processing, washing or storage of embryos were free of pathogenic organisms including pestiviruses. Media and solutions were sterilized by approved methods according to the IETS Manual and handled in such a manner as to ensure that sterility was maintained. Antibiotics as recommended in the OIE Code and IETS Manual, or a combination of antibiotics with equivalent activity, were added to collection, processing, washing and storage media.

    Name and concentration of antibiotics: _____

14. All straws were sealed, and clearly and permanently marked to identify the donor and the date(s) of freezing. If a code is used for this information, its decipher accompanies the consignment. The marking conforms to the international standards of the International Committee for Animal Recording (ICAR; www.icar.org) and the IETS.

15. Where embryos are removed from the transport containers for further processing, the date of transfer, approved embryo collection team, reason for transfer, and approved embryo collection team veterinarian must be recorded on the veterinary certificate.

    Date of transfer: _____

    Approved embryo collection team: _____

    Approved embryo collection team veterinarian: _____

    Reason for transfer: _____

16. The embryos for export were stored in the frozen state for at least 30 days after collection before shipment to New Zealand, and during this time, the donors and all animals in contact with them remained free from any diseases transmissible in embryos.

17. The embryos were only stored with germplasm that has been collected and processed in compliance with the OIE Code or semen collected and processed according to CSS standards. Containers were held until export in a storage place approved by APHIS.

18. The embryos were placed in transport containers filled with fresh (previously unused) liquid nitrogen. Transport containers are either new or empty and disinfected. For the transport container used to transport the embryos to New Zealand:

    Name of disinfectant: _____
    Disinfectant active chemical: _____
    Date of disinfection: _____

19. The transport container in which the embryos are to be transported to New Zealand was sealed by the approved embryo collection team veterinarian, using a tamper evident seal.

    Seal number: _____

Laboratory testing

20. All required laboratory testing was conducted at an approved laboratory and test results are documented in the table attached to this certificate.

21. Samples of embryos/oocytes, collection fluids, and washing fluids for laboratory testing were collected, processed, and stored in accordance with the recommendations in the OIE Code chapter on collection and processing of *in vivo* derived embryos of livestock.

22. Laboratory or other diagnostic tests were those prescribed for that disease by the OIE for use during international trade, or specifically approved by the Ministry of Primary Industries.

SPECIFIC REQUIREMENTS:

Bovine viral diarrhea type 2 (BVDV2) (delete section 23 or 24, as appropriate. and initial)
EITHER

23. Donors have been tested for BVDV including:
    a. prior to, or at the time of embryo collection for export to New Zealand, all donors were tested serologically for BVDV and for BVDV antigen using an OIE prescribed test; AND
    b. 21 to 40 days subsequent to embryo collection for export to New Zealand, seronegative donors were tested serologically for BVDV and for BVDV antigen using an OIE prescribed test.

    NB: Cattle that are not eligible as embryo donors for export to New Zealand are either:
    a. donors that are antigen-positive in initial testing; OR
    b. donors that seroconvert or are antigen-positive in the post-collection test.

OR

24. A pooled sample of embryos/oocytes, collection fluids and/or washing fluids from each embryo collection for export to New Zealand has been tested for BVDV2, by virus isolation (VI), with negative results.

Vesicular stomatitis (VS) (delete section 25, 26 or 27, as appropriate, and initial)
EITHER

25. Donors were resident in the United States, a country that is free from VS in accordance with the OIE Code;

OR

26. VS is officially notifiable in the United States, and no known cases have occurred within 100km of the embryo collection herd, where the donors were resident during embryo collection, during the period from 30 days prior to commencement until 30 days after conclusion of embryo collection for export to New Zealand;

OR

27. Donors were:
    a. resident for the 30 days prior to and during embryo collection in a herd where no case of VS was reported in that period; AND
    b. subjected to a serological test for VS, between 21 to 42 days after embryo collection for export to New Zealand, with negative results.

Bovine tuberculosis

28. Donors and other susceptible animals in the embryo collection herd showed no clinical signs of bovine tuberculosis during the 24 hours prior to embryo collection for export to New Zealand;

AND

29. Donors were:
    a. from an embryo collection herd that is officially free from bovine tuberculosis; AND
    b. subjected to a tuberculin test for bovine tuberculosis during the period between 30 days prior to 12 months after embryo collection for export to New Zealand, with negative results.

Mycoplasma bovis

30. Donors have never recorded a positive test for *Mycoplasma bovis*;

Q fever (delete section 32 or 33, as appropriate, and initial)

31. Donors have never been confirmed positive for Q fever;

AND EITHER

32. Donors were subjected to a CF or ELISA test for Q fever, on a sample collected between 21 and 120 days after each embryo collection for export to New Zealand, with negative results;

OR

33. Within the 6 month period before or after embryo collection for export to New Zealand, the embryo collection herd was tested for Q fever, with negative results, using a CF or ELISA test done on either the whole herd or a random sample of at least 60 animals (whichever is the lesser number); AND
    The embryo collection herd was isolated for the period between embryo collection and diagnostic sampling.

Approved Embryo Collection Team Veterinarian: Name and address (in capital letters): Date: Signature:	Official APHIS Veterinarian: Name and address (in capital letters): Date: Signature: Stamp:

Health Certificate No._____

(Valid only if USDA Veterinary Seal Appears Over the Certificate No.)

Test information (date format is dd-mm-yyyy)

		Bovine viral diarrhea (BVD2)			Q fever			Bovine tuberculosis			Vesicular stomatitis (if applicable)		
Donor registration name/number	Collection date	Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result	Test sampling date	Test type	Result