DEPARTMENT FOR ENVIRONMENT, FOOD AND RURAL AFFAIRS
SCOTTISH GOVERNMENT
WELSH GOVERNMENT
DEPARTMENT OF AGRICULTURE, ENVIRONMENT AND RURAL AFFAIRS - NORTHERN IRELAND

EXPORT OF IN-VIVO DERIVED OVINE AND CAPRINE EMBRYOS TO NEW ZEALAND

HEALTH CERTIFICATE **No:.....**

EXPORTING COUNTRY: **UNITED KINGDOM**

FOR COMPLETION BY: **OFFICIAL VETERINARIAN**

Part 1. Details of dispatched consignment

1.1 Name and address of consignor (exporter):

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1.2 Import permit number: .....

1.3 Competent authority:.....

1.4 Name and address of consignee (importer):

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1.5 Country of origin of embryos:.....

1.6 Place of origin of embryos (name and address of premises where embryos were collected):.....

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1.7 Approval number of Embryo Collection Team:.....

1.8 Embryo Collection Team Veterinarian

Name and qualifications

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Address

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....

1.9 The consignment of embryos is to be sent from (place of loading):

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....

1.10 Planned date of shipment:.....

1.11 Means of transportation (including registration number of vehicle, flight number of aircraft or name of ship:

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..

1.12 Expected border post:.....

Part 2: Detail of donors, germinal product, other

See attached supplementary sheet(s)

Part 3: Specific Requirements

I, ..., an Official Veterinarian, certify after due enquiry that the embryos described in Part 2 of this certificate satisfy the following requirements:

Eligibility

(1) The embryos are from * *Ovis aries* and/or * *Capra hircus*;

(2) The embryos are *in-vivo* derived, frozen, non-cloned and non-genetically modified;

Diagnostic testing, vaccination and treatment

(3) All required laboratory testing was conducted at a laboratory approved to conduct export testing by the Competent Authority of the United Kingdom or approved by the New Zealand Ministry of Primary Industries;

(4) All products and vaccinations administered to meet the specific disease requirements of this certificate were administered according to the manufacturer's instruction in a country approved to export ovine and caprine embryos to New Zealand. Vaccinations were either the final dose of a primary course or the recommended booster to complement the primary course.

Details of products and vaccines administered, including dates of treatment, are listed in Part 2 of this certificate;

Embryo collection centre team and flock/herd requirements

(5) At the time of collection of the embryos for export to New Zealand, the embryo collection team was approved by and registered with the Competent Authority of the exporting country;

(6) The Competent Authority had knowledge of and authority over the embryo collection flock/herd until completion of collection and testing specified in this certificate;

Donor and flock/herd health status

(7) The embryo donors were not situated in a flock/herd subject to veterinary restrictions for the identified risk organisms for at least 28 days before the first embryo collection until completion of the testing of the donors as required by this certificate;

(8) Where a specific requirement for a risk organism is met by pre-collection testing, donors were isolated from other sheep or goats not of an equivalent tested health status from the time of the pre-collection test until completion of collection of embryos for export to New Zealand;

(9) On the day(s) of embryo collection, the approved embryo collection team veterinarian was responsible for monitoring the health status of each donor and recording that the donor was free from clinical evidence of infectious disease transmissible in embryos;

Embryo collection, processing, storage and transport

(10) The embryos were collected and processed under the supervision of an approved embryo collection team veterinarian and in accordance with the protocols detailed in the current edition of the Manual of the International Embryo Transfer Society (IETS) on collection and processing of *in vivo* derived embryos of livestock, unless indicated otherwise in this certificate;

(11) The embryos had an intact zona pellucida and were free of adherent material after the final wash when examined over its entire surface at not less than 50X magnification. Any micromanipulation that caused a breach of the zona pellucida was performed according to the procedures described in the OIE Code and the International Embryo Transfer Society (IETS) Manual;

(12) All biological products of animal origin used in the media and solutions for collection, processing, washing or storage of the embryos were free from pathogenic organisms including pestiviruses;

(13) Media and solutions were either sterilised by approved methods according to the IETS Manual or commercially prepared sterile media were used.

These were handled in such a manner as to ensure that sterility was maintained;

(14) The collection, processing, washing and storage media have been treated by the addition of antibiotics effective against *Leptospira* serovars, as recommended in the OIE Code;

(15) None of the cryogenic or cooling agents have been previously used in association with any other product of animal origin;

(16) The embryos are sealed in straws, which are clearly and permanently marked to identify the donor and the date(s) of collection.

*A code is used for this information, as follows:

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.....

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The marking is in accordance with the OIE Code and conforms to the international standards of the International Committee for Animal Recording (ICAR);

(17) The embryos were only stored with semen or embryos that were collected and processed according to the OIE Code. Storage containers were held until export in a storage place approved by the Competent Authority of the exporting country;

*(18) Embryos were transferred from one transport container to another for further processing. Transfer date, location and reason:

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(19) The embryos were placed in a transport container which *was new or *had been cleaned and disinfected. Name of disinfectant (active chemical) and date:

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(20) The transport container in which the embryos will be transported to New Zealand was sealed by either the embryo collection team veterinarian or an Official Veterinarian, using tamper-evident seals.

Seal number:.....

*(21) The embryos in this consignment originate from a different country, namely.....(insert name of country). This country was approved at the time of collection, and is still approved, to export ovine and caprine embryos to New Zealand.

The embryos were accompanied by:

(a) A declaration from the Competent Authority of the Country linking the embryos from the country of origin to the embryos being exported to New Zealand and confirming that the embryos have been stored as required by the certificate at a facility approved by the Competent Authority; AND

*(b)(i) The veterinary certificate certified by the country of origin to export to New Zealand requirements; OR

*(b)(ii) A letter from the country of origin's Competent Authority indicating that the embryos meet New Zealand's current import requirements;

CONTINUED ON 7855CON

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HEALTH CERTIFICATE (continuation) **No: .....**

EXPORTING COUNTRY: **UNITED KINGDOM**

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Part 3: Specific Requirements (continued from 7855EHC)

SPECIFIC REQUIREMENTS FOR IDENTIFIED RISK ORGANISMS

(22) Bluetongue virus (BTV)

The embryos were obtained from donor(s) which comply with at least one of the following conditions:

*(a) they were kept in a BTV free country or zone in accordance with the requirements of the OIE Code for at least the 60 days prior to, and at the time of, collection of the embryos; OR

*(b) they were subjected to an ELISA according to the OIE Terrestrial Manual to detect antibodies to the BTV group, with negative results, carried out on blood samples collected between 28 and 60 days after collection; OR

*(c) they were subjected, with negative results, to an agent identification test for BTV according to the OIE Terrestrial Manual carried out on a blood sample taken on the day of collection of the embryos; OR

*(d) they were vaccinated with a vaccine listed in the MPI-STD-TVTL against all known BTV serotypes in the United Kingdom, no less than 2 months and no more than one year before collection;

(23) Foot and mouth disease (FMD)

*(a) The donors were kept for at least 90 days prior to and during collection of the embryos in this consignment in a FMD-free country or zone without vaccination, in accordance with the OIE Code, and showed no clinical signs of FMD during the 30 days after collection; OR
*(b) The donors were kept in a flock/herd/facility where no animal was added in the 30 days before collection; and

(i) For the 30 days after collection neither the donors nor any other animal where the donors were kept showed clinical signs of FMD; and

(ii) FMD has not occurred within a 10 kilometre radius of the flock/herd/facility for the 30 days before and after collection; and either

*1. Donors have been vaccinated at least twice with the last vaccination not less than one month and not more than six months prior to collection, unless protective immunity has been demonstrated for more than six months; or

*2. Donors were subjected, not less than 21 days after collection of the embryos, to a Virus Neutralisation Test (VNT) or enzyme-linked immunosorbent assay (ELISA) for antibodies against FMDV, with negative results; and

*(iii) If the donor was vaccinated within the 12 months prior to collection, the non-viable embryos/washing/flushing fluid was subjected, with negative results, to a virus isolation test for evidence of FMDV;

(24) Maedi-visna virus (MV)

(a) The donors were tested for MV with negative results using either *agar gel immunodiffusion test (AGIDT) or *enzyme-linked immunosorbent assay (ELISA), during the 21 day period prior to embryo collection; and

(i) All embryos were washed in trypsin, according to the recommendations of IETS;

(25) Peste des petits ruminants virus (PPR)

The donors were resident in a PPR-free country or zone in accordance with the OIE Code for at least 21 days prior to and during collection of the embryos in this consignment;

(26) Rift Valley fever virus (RVF)

The donors were resident in a RVF-free country or zone in accordance with the OIE Code for at least 30 days prior to and during collection of the embryos in this consignment;

(27) Capripox virus (sheep and goat pox)

The donors were resident in a sheep and goat pox-free country in accordance with the OIE Code for at least 21 days prior to and during collection of the embryos in this consignment;

(28) Wesselsbron disease virus (Wesselsbron disease)

The donors were resident in a country recognised by the Competent Authority as free from circulating Wesselsbron disease virus for at least 21 days prior to and during collection of the embryos in this consignment;

(29) *Brucella melitensis* (caprine and ovine brucellosis)

*(a) The donors were resident in a country, zone, or herd that is officially free from caprine and ovine brucellosis in accordance with the OIE Code and the donors were not vaccinated against Brucellosis in the past 3 years; OR

*(b) The donor animals were not vaccinated against infection with *Brucella* in the past three years and were resident in a flock/herd that is free from caprine and ovine brucellosis in accordance with the OIE Code and were tested for caprine and ovine brucellosis (*Brucella melitensis*), with negative results, using either complement fixation test (CFT) or enzyme-linked immunosorbent assay (ELISA) within 30 days prior to the first collection of embryos in this consignment;

***(30) *Mycoplasma capricolum* subsp. *Capripneumoniae* (contagious caprine pleuropneumonia – CCPP)**

For goats only: The donors were resident in a country that is free from CCPP in accordance with the OIE Code;

(31) *Mycoplasma agalactiae* (contagious agalactia)

The donors were resident in a country recognised by the Competent Authority as free from contagious agalactia for at least 6 months prior to and during collection of the embryos in this consignment;

***(32) *Mycobacterium caprae* and *Mycobacterium bovis* (tuberculosis)**

For goats only:

(a) During the 28 days prior to collection of the embryos in this consignment, there were no signs of tuberculosis in the flock/herd and the donors were subjected to a comparative intradermal tuberculin test using avian and bovine purified protein derivative (PPD) tuberculins, with negative results according to the Department's standard interpretation, AND

(b) Donors were kept in herds free from bovine tuberculosis and tested annually with negative results with the test described in (i);

(33) *Chlamydia abortus* (enzootic abortion of ewes – EAE)

*****(a) The donors were resident in a flock/herd that is free from EAE in accordance with the OIE Code for at least the 2 years prior to embryo collection and were not in contact with any animals of lower health status during that period of time; OR

*****(b) The donors have been resident since birth, or for at least the two years prior to embryo collection, in a flock/herd where no EAE has been diagnosed and: either

*(i) The donors were tested for EAE, with negative results, using the complement fixation test (CFT); the samples were collected at least 21 days after the final collection of the embryos in this consignment; or
*(ii) Embryos/oocytes or collection/washing fluids were subjected to a validated PCR test from the end of each collection period (60 days or less); OR

*(c) The semen used to fertilise the embryo satisfies New Zealand's import requirements for semen from sheep and goats; and
*(i) The donor is not known to have ever aborted a foetus during the last month of gestation, had a stillbirth, or an abnormally weak neonate; and

*(ii) The donor has been resident since birth, or for at least the two years prior to collection, only in flocks/herds that either:
*1. are free in accordance with the Code; or
*2. have no history of late gestation abortions for the past 2 years and all female animals introduced during that time have tested seronegative for EAE after joining the herd/flock; or
*3. tested placentae, uterine discharges, or the foetus/neonate, from every late gestation abortion/stillbirth/weak neonate, for EAE as per the OIE Manual, during the past 2 years, with negative results; or
*4. conducted serological screening[1] of ewes for the 2 years before collection, testing at the time of

abortion/parturition and between 2 and 4 weeks later, and there have been no rises in titre.

[1]Screening must be randomised and representative of the herd/flock. The sample size selected must be sufficiently large to give 95% confidence of detecting infection.

(34) *Coxiella burnetii* (Q fever)

***(a)** Donors

(i) prior to 1st vaccination, only resided in herds/flocks where, for the previous 4 years, the abortion rate was:

*1. 2% or under; or

*2. investigated and Q fever was never diagnosed; and

*(ii) recorded a negative *ELISA or *IFA at the time of vaccination; and

*(iii) were vaccinated with an inactivated whole phase 1 vaccine, as per the OIE Manual. That vaccination, or a booster, was administered within the 12 months before collection; and since vaccination either

*1. The donor only resided in flocks where there was no evidence of Q fever for at least the previous 4 years; or

*2. Every flock where the donor resided for the past 2 years, PCR tested uterine discharges or foetuses from all late gestation abortion/stillbirth/weak neonate for Q fever, as per the Manual, with negative results; OR

*(b) The donors have never been confirmed positive for Q fever and either

*(i) The donors were tested for Q fever, with negative results, using an enzyme-linked immunosorbent assay (ELISA), on a sample collected between 21 and 120 days after collection for export to New Zealand; or

*(ii) Embryos/oocytes or collection/washing fluids were subjected to a validated PCR test from the end of each collection period (60 days or less).

***(35) Scrapie**

For goats only:

(a) The donors were resident in a collection herd that has been maintained free from scrapie from commencement until conclusion of embryo collection, in accordance with the OIE Code recommendations for a scrapie-free establishment; OR

(b) The donors were permanently identified to enable trace back to their establishment of origin and were kept in establishments since birth in which no case of scrapie was confirmed during their residency.

*** Delete as appropriate and initial**

Official Stamp **Signed ..... RCVS Official Veterinarian**

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Name in block letters

Date ..... **Address** .....

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