

**VETERINARY HEALTH CERTIFICATE
 EXPORT OF OVINE EMBRYOS TO NEW ZEALAND**

Part I

1. Consignor (Exporter): Name: Address:	2. Certificate reference number:								
	3. Veterinary Authority: <p align="center">CANADIAN FOOD INSPECTION AGENCY</p>								
	4. Import permit number:								
5. Consignee (Importer): Name: Address:	6. Country of origin: CANADA								
	7. Country of destination: NEW ZEALAND								
8. Place of shipment:	9. Date of departure:								
10. Description of commodity: OVINE EMBRYOS	11. Total quantity:								
12. Identification of container(s) and seal number: <table border="0" style="width:100%"> <tr> <td style="width:50%">Container</td> <td style="width:50%">Seal Number</td> </tr> <tr> <td>_____</td> <td>_____</td> </tr> <tr> <td>_____</td> <td>_____</td> </tr> <tr> <td>_____</td> <td>_____</td> </tr> </table>	Container	Seal Number	_____	_____	_____	_____	_____	_____	13. Antibiotics (and their concentration) added to germplasm:
Container	Seal Number								
_____	_____								
_____	_____								
_____	_____								
14. Transport container : New or Disinfected (delete as appropriate and initial) If disinfection: Disinfectant used: _____ Active chemical: _____ Date of disinfection (yyyy-mm-dd): _____									

**Part II
HEALTH CERTIFICATION**

I,....., a veterinarian authorised by the Competent Authority certify, after due enquiry that the donor animals and embryos described in this certificate satisfy the following requirements:

Eligibility

- The embryos are from *Ovis aries*
- The embryos are in vivo derived, frozen, non-cloned, and non-genetically modified.

Laboratory testing

- All required laboratory testing was conducted in a laboratory of the Competent Authority or a laboratory approved by the Competent Authority.
- Laboratory tests or other diagnostic tests are those prescribed for this disease by the OIE Manual, or specifically approved by New Zealand (Ministry for Primary Industries, MPI).
- An endorsed tabulated summary, including test date, type, and results for each donor, is included in this veterinary health certificate.

Embryo collection team and flock approval requirements

- At the time of collection of embryos for export to New Zealand, the embryo collection team was approved by and registered with the Competent Authority.
- The Competent Authority has knowledge of and authority over the embryo collection flock until completion of collection and testing specified in this veterinary health certificate.

Donor and flock health status

- Embryo donors were not situated in a flock subject to veterinary restrictions for the identified risk organisms, for at least 28 days before the first embryo collection until completion of the testing of the donors as stated on this veterinary health certificate.

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9. Where a specific requirement for a risk organism is met by pre-collection testing, donors were isolated from other sheep not of an equivalent tested health status, from the time of the pre-collection test until completion of collection of embryos for export to New Zealand.
10. On the day(s) of embryo collection, the approved embryo collection team veterinarian was responsible for monitoring the health status of each donor and recording that the donor was free from clinical evidence of infectious diseases transmissible in embryos.

Embryo collection, processing, storage and transport

11. Embryos were collected and processed under the supervision of an approved embryo collection team veterinarian and in accordance with the recommendations in the OIE Code chapters on collection and processing of *in vivo* derived embryos of livestock.
12. To manage *Leptospira* serovars, antibiotics, as recommended in the IETS Manual, were added to collection media. All processing, washing and storage media were sterile. The names of antibiotics added and their concentration are stated on this veterinary certificate.
13. Embryos had an intact zona pellucida and were free of adherent material after the final wash when examined over its entire surface at not less than 50X magnification. Any micro-manipulation that caused a breach of the zona pellucida, was performed according to the procedures described in the Code and IETS Manual.
14. All biological products of animal origin used in the media and solutions for collection, processing, washing or storage of embryos were free from pathogenic organisms including pestiviruses.
15. Media and solutions were either sterilised by approved methods according to the IETS Manual or commercially prepared sterile media were used. These were handled in such a manner as to ensure that sterility was maintained.
16. None of the cryogenic or cooling agent has been previously used in association with any other product of animal origin.
17. All straws are sealed, and clearly and permanently marked to identify the donor and the date of collection. The marking should, in accordance with the OIE Code, conform to the international standards of the IETS.
18. Embryos were only stored with germplasm that were collected and processed according to the *Code*. Containers were held until export in a storage place approved by the Competent Authority.
19. The embryos were placed in a container which is sanitised and free of contamination. Disinfectant (active chemical) and date (delete and initial if the container was new): _____
20. The transport container, in which the embryos are transported to New Zealand, was sealed by an official veterinarian, using tamper evident seals. The seal number is recorded on this veterinary health certificate.

SPECIFIC REQUIREMENTS FOR IDENTIFIED RISK ORGANISMS:

Bluetongue virus –BTV (bluetongue) (strike out and initial statements that do not apply to the shipment)

21. The embryos were collected prior to September 3, 2015, or between January 1st and May 15 (seasonally free period),
or
22. The embryos were collected after September 3, 2015 and between May 16 and December 31, and donors were subjected to an ELISA test for BTV in accordance with the OIE Manual, with negative results, on blood samples taken on the day of collection for this consignment.

Crimean Congo haemorrhagic fever (CCHF)

23. CCHF has never been reported in animals in Canada.

Foot and mouth disease (FMD)

24. Donors were resident for at least the 3 months before embryo collection in Canada or another country approved to export sheep and goat embryo to New Zealand and neither the donors nor any other animal in the collection flock showed clinical signs of FMD on the day of the embryo collection for New Zealand and for the following 30 days. Canada is officially recognized free of FMD without vaccination by the OIE.

Maedi-visna virus (MV)

25. Donors were subjected to an ELISA test for MV in accordance with the OIE Manual, with negative results, at least 21 days after entering the collection flock and at least annually thereafter while in the collection flock
and
26. All embryos were washed in trypsin, according to the recommendations of IETS.

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Peste des petits ruminants (PPR)

27. Donors were resident in a PPR free country in accordance with the Code for at least 21 days prior to and during embryo collection. Canada is officially recognized free of PPR by the OIE.

Rift Valley fever (RVF)

28. Donors were resident in a RVF free country in accordance with the Code for at least the 30 days prior to embryo collection. RVF has never been reported in Canada.

Capripox virus (sheep and goat pox)

29. Donors were resident in a sheep and goat pox free country in accordance with the Code for at least 21 days prior to collection. Sheep and goat pox has never been reported in Canada.

Wesselsbron disease virus (Wesselsbron disease)

30. Donors were resident in a country recognised by the competent authority as free from circulating Wesselsbron disease virus for at least 21 days prior to collection. Wesselsbron disease has never been reported in Canada.

Brucella melitensis (ovine brucellosis)

31. Donors were resident in a country that is officially free from ovine brucellosis in accordance with the Code. *Brucella melitensis* has never been reported in Canada.

Mycoplasma agalactiae (contagious agalactia)

32. Donors were resident in a country that has been recognised by the competent authority as free from contagious agalactia for at least the 6 months prior to collection.

Chlamydia abortus (enzootic abortion of ewes – EAE)

33. Donor animals showed no clinical sign on the day of the embryo collection,
and either
 34. have been kept in flocks or centre free from EAE in accordance with the OIE Code for the two years prior to collection, and have not been in contact with animals of a lower health status;
or
 35. have remained since birth, or for the two years prior to collection, in establishments where no EAE has been diagnosed and were subjected to a CF test for EAE in accordance with the OIE Manual, with negative results as per the OIE recommendation, between two and four weeks after collection of the embryos.

Coxiella burnetii (Q fever)

36. Donors have never been confirmed positive for Q fever,
and
 37. were subjected to an ELISA test for Q fever in accordance with the OIE Manual, with negative results, on a sample collected between 21 and 120 days after each embryo collection for export to New Zealand.

 Embryo collection team Veterinarian

 ET Code

 Signature of Team Veterinarian

 Date (yyyy-mm-dd)

 Signature of Official Veterinarian
 Canadian Food Inspection Agency

 Name of Official Veterinarian (in block letters)
 Address: _____

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Note: Official Veterinarian signature and Official Stamp must be apply on each page

REFERENCE NUMBER: _____

Table I: Female Donor Information

Name	Registration Number	Donor identification	Breed	Date of Birth (yyyy-mm-dd)	Country of birth	Name of Owner	Address of Owner

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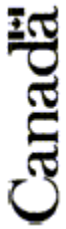
REFERENCE NUMBER: _____

Table II: Embryo Information (date format is yyyy-mm-dd)

Female donor identification	Date(s) of Collection (yyyy-mm-dd)	Straw identification	Number of straws	Number of embryos/ straw	Name and address of embryo collection flock	Male donor identification	Date of semen collection or date of natural mating
		Straws are identified with donor registration number, donor name, practitioner code, and freezing date					

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REFERENCE NUMBER: _____

Table III: Test Information (date format is yyyy-mm-dd). When a test does not apply, insert N/A (not applicable).

Female donor Identification	Date(s) of collection	Bluetongue, as per art. 22			Maedi-visna, as per art. 25			Enzootic Abortion in Ewes, as per art. 35			Q fever, as per art. 37		
		Sampling date	Test type	Result	Sampling date	Test type	Results	Sampling date	Test type	Results	Sampling date	Test type	Results

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